

Chemical and physical characteristics of the principal mycotoxins

Summary

This chapter provides information about the chemical and physical properties of the mycotoxins considered in this book: aflatoxins; fumonisins; ochratoxin A; trichothecenes, especially deoxynivalenol and nivalenol; zearalenone; and ergot alkaloids. This information about structures reveals the chemical diversity of mycotoxins, which is relevant to the wide range of toxicological effects in animals and humans discussed later in the book.

1. Aflatoxins

1.1 Formulae and structures

Aflatoxin B₁. Chemical Abstracts (CA) name: (6aR,9aS)-2,3,6a,9a-tetrahydro-4-methoxycyclopenta[c]furo-(3',2':4,5)-furo[2,3-*h*][l]benzopyran-1,11-dione. Chemical Abstracts Service (CAS) regis-

try number: 1162-65-8. Molecular formula: C₁₇H₁₂O₆. Molecular weight: 312.3.

Aflatoxin B₂. CA name: (6aR,9aS)-2,3,6a,8,9,9a-hexahydro-4-methoxycyclopenta[c]furo(3',2':4,5)furo[2,3-*h*][l]benzopyran-1,11-dione. CAS registry number: 7220-81-7. Molecular formula: C₁₇H₁₄O₆. Molecular weight: 314.3.

Aflatoxin G₁. CA name: (7aR,10aS)-3,4,7a,10a-tetrahydro-5-methoxy-1*H*,12*H*-furo-(3',2':4,5)furo[2,3-*h*]pyrano[3,4-*c*][l]benzopyran-1,12-dione. CAS registry number: 1165-39-5. Molecular formula: C₁₇H₁₂O₇. Molecular weight: 328.3.

Aflatoxin G₂. CA name: (7aR,10aS)-3,4,7a,9,10,10a-hexahydro-5-methoxy-1*H*,12*H*-furo-(3',2':4,5)furo[2,3-*h*]pyrano[3,4-*c*][l]benzopyran-1,12-dione. CAS registry number: 7241-98-7. Molecular formula: C₁₇H₁₄O₇. Molecular weight: 330.3.

Aflatoxin M₁. CA name: (6aR,9aR)-2,3,6a,9a-tetrahydro-9a-hydroxy-4-methoxycyclopenta[c]furo-(3',2':4,5)-furo[2,3-*h*][l]benzopyran-1,11-dione. CAS registry number: 6795-23-9. Molecular formula: C₁₇H₁₂O₇. Molecular weight: 328.3.

Structures of aflatoxins are shown in Fig. 2.1.

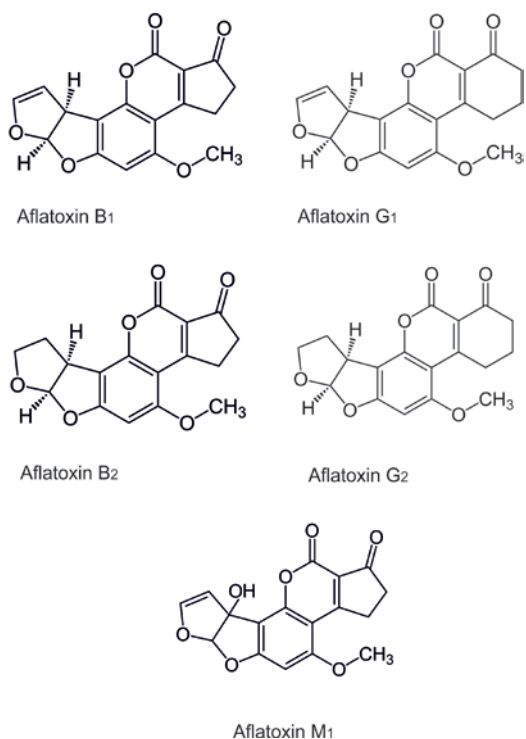
1.2 Physical data

Descriptions. Colourless to pale-yellow crystals. Fluorescence in ultraviolet (UV) light: aflatoxins B₁ and B₂, blue; aflatoxins G₁ and G₂, green; aflatoxin M₁, blue-violet.

Melting-points. See Table 2.1.

Spectral properties. For UV absorption, see Table 2.1. Fluorescence excitation and emission data are not listed in Table 2.1 because they depend on the type of instrument, the solvent, and the supporting media

Fig. 2.1. Structures of aflatoxins



used. For those data, see Wogan (1966), Robertson and Pons (1968), Kiermeier and Kroczeck (1974), and Uwaifo *et al.* (1977).

For mass and nuclear magnetic resonance (NMR) spectral data, see Bycroft *et al.* (1970), Stubblefield *et al.* (1970), and Cole and Schweikert (2003).

Specific rotation. $[\alpha]_D$ in chloroform, -558° (aflatoxin B₁), -430° (aflatoxin B₂), -556° (aflatoxin G₁), -473° (aflatoxin G₂); $[\alpha]_D$ in dimethylformamide, -280° (aflatoxin M₁) (Cole and Schweikert, 2003).

1.3 Chemical data

Solubility. Insoluble in non-polar solvents. Slightly soluble in water (10–20 $\mu\text{g/mL}$). Freely soluble in moderately polar organic solvents (e.g. chloroform, methanol), especially in dimethyl sulfoxide (Cole and Cox, 1981; O’Neil *et al.*, 2001).

Stability. Unstable to UV light in the presence of oxygen. Unstable to extremes of pH (< 3 or > 10). Unstable in the presence of oxidizing agents (Castegnaro *et al.*, 1980, 1991).

Reactivity. Under alkaline conditions, the lactone ring opens and the aflatoxins are apparently absent. However, the reaction is reversible upon acidification.

Ammoniation at high temperature and high pressure opens the lactone ring and results in decarboxylation. This reaction is not reversible.

2. Fumonisin

2.1 Formulae and structures

Fumonisin B₁. CA name: 1,2,3-propanetricarboxylic acid, 1,1'-[1-(12-amino-4,9,11-trihydroxy-2-methyltridecyl)-2-(1-methylpentyl)-1,2-ethanediyl] ester. CAS registry number: 116355-83-0. Molecular formula: C₃₄H₅₉NO₁₅. Molecular weight: 721.

Fumonisin B₂. CA name: 1,2,3-propanetricarboxylic acid, 1,1'-[1-(12-amino-9,11-dihydroxy-2-methyltridecyl)-2-(1-methylpentyl)-1,2-ethanediyl] ester. CAS registry number: 116355-84-1. Molecular formula: C₃₄H₅₉NO₁₄. Molecular weight: 705.

Structures of fumonisins are shown in Fig. 2.2.

2.2 Physical data

Unless otherwise noted, data are from WHO (2000).

Description. White hygroscopic powder.

Melting-point. Not known (compounds have not been crystallized).

Spectral properties. For mass and NMR spectral data, see Bezuidenhout *et al.* (1988), Laurent *et al.* (1989), Plattner *et al.* (1990), Savard and Blackwell (1994), and Cole *et al.* (2003a).

2.3 Chemical data

Solubility. Soluble in methanol, in acetonitrile–water, and in water (at least 20 g/L) (NTP, 2001).

Stability. Stable in acetonitrile–water (1:1) at 25 °C. Unstable in methanol at 25 °C, forming monomethyl and dimethyl esters (Gelderblom *et al.*, 1992; Visconti *et al.*, 1994). Stable in methanol at -18°C (Visconti *et al.*, 1994). Stable in buffer solutions over the pH range 4.8–9 at 78 °C (Howard *et al.*, 1998).

Octanol–water partition coefficient for fumonisin B₁. $\log P = 1.84$ (Norred *et al.*, 1997).

3. Ochratoxin A

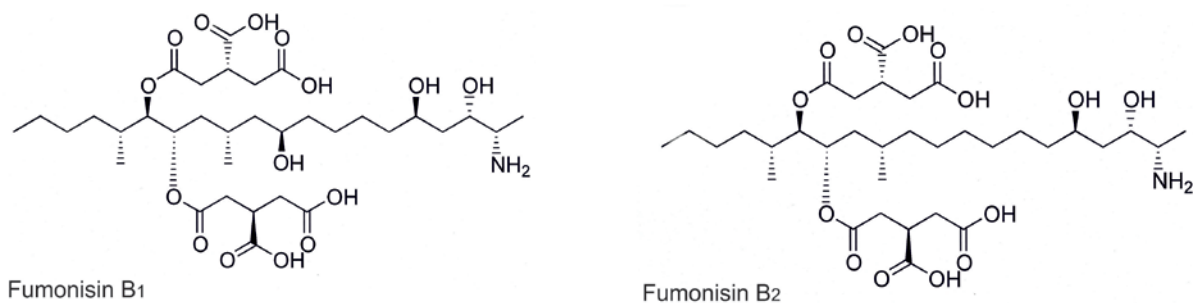
3.1 Formula and structure

Ochratoxin A. CA name: *N*-[(5-chloro-3,4-dihydro-8-hydroxy-3-methyl-1-oxo-1*H*-2-benzopyran-7-yl)carbonyl]-L-phenylalanine. CAS registry number:

Table 2.1. Melting-points and ultraviolet absorption of aflatoxins

Aflatoxin	Melting-point (°C)	Ultraviolet absorption	
		λ_{\max} (nm)	ϵ (L·mol ⁻¹ ·cm ⁻¹) × 10 ⁻³
B ₁	268–269 (decomposition) (crystals from CHCl ₃)	223	25.6
		265	13.4
		362	21.8
B ₂	286–289 (decomposition) (crystals from CHCl ₃ –pentane)	265	11.7
		363	23.4
G ₁	244–246 (decomposition) (crystals from CHCl ₃ –methanol)	243	11.5
		257	9.9
		264	10.0
		362	16.1
G ₂	237–240 (decomposition) (crystals from ethyl acetate)	265	9.7
		363	21.0
M ₁	299 (decomposition) (crystals from methanol)	226	23.1
		265	11.6
		357	19.0

Data from O'Neil *et al.* (2001).

Fig. 2.2. Structures of fumonisins

303-47-9. Molecular formula: $C_{20}H_{18}ClNO_6$. Molecular weight: 403.8.

The structure of ochratoxin A (OTA) is shown in Fig. 2.3.

3.2 Physical data

Description. White odourless crystalline solid (Pohland *et al.*, 1982). Intensely fluorescent in UV light, emitting green and blue fluorescence in acid and alkaline solutions, respectively, due to two different forms, i.e. closed or open lactone ring, respectively.

Melting-point. 159 °C when recrystallized from benzene–hexane (Natori *et al.*, 1970); 169 °C when recrystallized from xylene (Van der Merwe *et al.*, 1965a, 1965b); 168–173 °C after drying for 1 hour at 60 °C (Pohland *et al.*, 1982).

Specific rotation. $[\alpha]_D^{20} -118^\circ$ ($c = 1.1$ mmol/L in chloroform) (Van der Merwe *et al.*, 1965a, 1965b); $[\alpha]_D^{21} -46.8^\circ$ ($c = 2.65$ mmol/L in chloroform) (Pohland *et al.*, 1982).

UV spectrum. At λ_{max} of 214, 282, and 332 nm, extinction coefficients of 37.2×10^{-3} , 0.89×10^{-3} , and $63.3 \times 10^{-3} \text{ L}\cdot\text{mol}^{-1}\cdot\text{cm}^{-1}$, respectively, have been reported (Cole and Cox, 1981).

Other spectral properties. For infrared (IR) spectra, see Van der Merwe *et al.* (1965a, 1965b), Steyn and Holzapfel (1967), and Pohland *et al.* (1982). For NMR spectra, see Pohland *et al.* (1982) and Cole *et al.* (2003b). For mass spectra, see Pohland *et al.* (1982) and Cole *et al.* (2003b).

3.3 Chemical data

Solubility. Moderately soluble in polar organic solvents (e.g. chloroform, ethanol, methanol).

Stability. OTA is partially degraded under normal cooking conditions (Müller, 1983). The stability of OTA to heating conditions depends on the water activity of the

medium (Subirade, 1996; Van der Stegen *et al.*, 2001).

Reactivity. The lactone ring opens under alkaline conditions, but the reaction is reversible. Solutions of OTA are completely degraded by treatment with an excess of sodium hypochlorite.

4. Deoxynivalenol

4.1 Formula and structure

Deoxynivalenol. CA name: 12,13-epoxy-3,7,15-trihydroxy-(3 α ,7 α)-trichothec-9-en-8-one. CAS registry number: 51481-10-8. Molecular formula: $C_{15}H_{20}O_6$. Molecular weight: 296.32.

The structure of deoxynivalenol (DON) is shown in Fig. 2.4.

4.2 Physical data

Description. White needles.

Melting-point. 151–153 °C.

Specific rotation. $[\alpha]_D^{20} +6.35^\circ$ ($c = 0.07$ mmol/L in ethanol).

Spectral properties. IR, UV, NMR, and mass spectral data have been reported (Cole and Cox, 1981; Cole *et al.*, 2003c).

4.3 Chemical data

Solubility. Soluble in chloroform, ethanol, methanol, and ethyl acetate.

Stability. Autoclaving creamed maize reduced DON content by only 12% (Wolf-Hall *et al.*, 1999). At pH 4.0, DON appeared to be very stable, showing no destruction at 100 °C or 120 °C and only partial destruction at 170 °C after 60 minutes. At pH 7.0, DON was still stable but showed more destruction at 170 °C after 15 minutes. At pH 10.0, DON was partially destroyed at 100 °C after 60 minutes and was totally destroyed at 120 °C after 30 minutes and at 170 °C after 15 minutes (Wolf and Bullerman, 1998).

When DON was gamma-irradiated on maize, breakdown of DON began only after irradiation to 20 kGy, and 80–90% of the DON remained after irradiation to 50 kGy (O'Neill *et al.*, 1993).

No significant decomposition of DON was observed when stored in ethyl acetate for 24 months at 25 °C or 3 months at 40 °C (Widestrand and Pettersson, 2001). DON was relatively stable in buffer solutions over the pH range 1–10 (Lauren and Smith, 2001).

5. Nivalenol

5.1 Formula and structure

Nivalenol. CA name: 12,13-epoxy-3,4,7,15-tetrahydroxy-(3 α ,4 β ,7 α)-trichothec-9-en-8-one. CAS registry number: 23282-20-4. Molecular formula: $C_{15}H_{20}O_7$. Molecular weight: 312.32.

The structure of nivalenol (NIV) is shown in Fig. 2.4.

5.2 Physical data

Description. White crystals.

Melting-point. 222–223 °C (with decomposition, after drying in the presence of P_2O_5 at reduced pressure).

Specific rotation. $[\alpha]_D^{20} +21.54^\circ$ ($c = 1.3$ mmol/L in ethanol).

Spectral properties. IR, UV, NMR, and mass spectral data have been reported (Cole and Cox, 1981; Brumley *et al.*, 1982; Cole *et al.*, 2003c).

5.3 Chemical data

Solubility. Soluble in chloroform, ethanol, methanol, and ethyl acetate; slightly soluble in water; soluble in polar organic solvents (Budavari, 1989).

Stability. No significant decomposition of NIV was observed when stored in ethyl acetate for 24 months at 25 °C or for 3 months at 40 °C. A significant decrease of NIV stored

Fig. 2.3. Structure of ochratoxin A

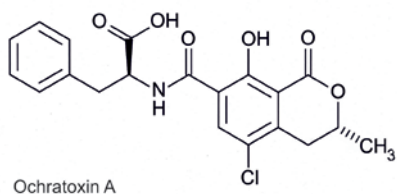


Fig. 2.4. Structures of major trichothecenes

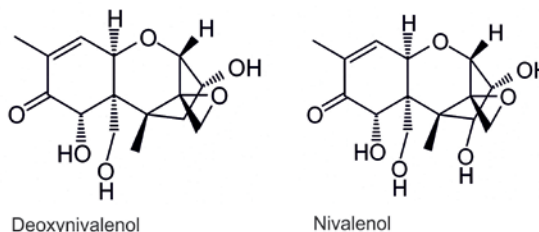


Fig. 2.5. Structure of zearalenone

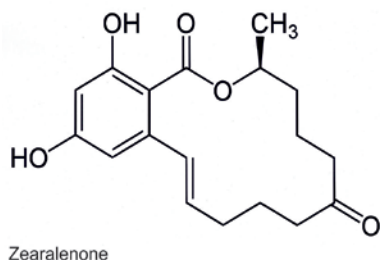
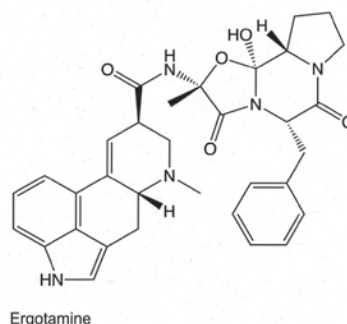


Fig. 2.6. Structure of ergotamine



as a thin film was observed after 9 months at 25 °C (Widestrand and Petterson, 2001). NIV is relatively stable in buffer solutions over the pH range 1–10 (Lauren and Smith, 2001).

6. Zearalenone

6.1 Formula and structure

Zearalenone. CA name: 3,4,5,6,9,10-hexahydro-14,16-dihydroxy-3-methyl-1*H*-2-benzoxacyclotetradecin-1,7(8*H*)-dione. CAS registry number: 17924-92-4. Molecular formula: C₁₈H₂₂O₅. Molecular weight: 318.4.

The structure of zearalenone (ZEA) is shown in Fig. 2.5.

6.2 Physical data

Description. White crystals.

Melting-point. 164–165 °C.

Specific rotation. $[\alpha]_D^{25}$ –170.5° (c = 1.0 mmol/L in methanol); $[\alpha]_D^{21}$ –189° (c = 3.14 mmol/L in chloroform).

Spectral properties. IR, UV, proton NMR, and mass spectral data have been reported (Cole and Cox, 1981). The molar absorptivities of ZEA in acetonitrile at 236, 274, and 314 nm were established, and a common reference wavelength of 274 nm with molar absorptivity of $12\,623 \pm 111 \text{ L}\cdot\text{mol}^{-1}\cdot\text{cm}^{-1}$ was recommended for ZEA in acetonitrile (Josephs *et al.*, 2003).

6.3 Chemical data

Solubility. Solubilities at 25 °C in percent by weight are: water, 0.002; *n*-hexane, 0.05; benzene, 1.13; acetonitrile, 8.6; dichloromethane, 17.5; methanol, 18; ethanol, 24; and acetone, 58 (Hidy *et al.*, 1977).

Stability. ZEA was stable when heated at 120 °C; 29% decomposed when heated at 150 °C for 60 minutes and 69% when heated at 200 °C for 60 minutes (Kuiper-Goodman *et al.*, 1987). Stable to hydrolysis in neutral or acid buffer solutions (Müller, 1983).

Less than 23% of ZEA was lost when heated in aqueous buffer solution to 125 °C for 60 minutes, but 34–68% was lost after 60 minutes at 150 °C, depending on the pH of the buffer. More than 92% was lost after 60 minutes at 175 °C, and complete loss was observed in < 30 minutes at 225 °C, regardless of pH. ZEA was most stable at pH 7, and the greatest losses occurred above 175 °C (Ryu *et al.*, 2003).

Extrusion cooking of maize grits resulted in significant reductions of ZEA with either mixing screws or non-mixing screws, but use of mixing screws was somewhat more effective (66–83% reduction) overall than non-mixing screws (65–77%). Greater reduction of ZEA content was observed at either 120 °C or 140 °C than at 160 °C (Ryu *et al.*, 1999).

ZEA content was not reduced by heating at 110 °C for 12 days after treatment with a sodium bicarbonate solution (Lauren and Smith, 2001).

7. Ergot alkaloids

Ergots, the sclerotia produced by *Claviceps purpurea* and related species, contain a remarkable variety of compounds, which can be divided into three groups: derivatives of lysergic acid, derivatives of isolysergic acid, and clavines. The most important of these is ergotamine.

7.1 Formula and structure

Ergotamine. CA name: 12'-hydroxy-2'-methyl-5'-(phenylmethyl)-ergotaman-

3',6',18-trione. CAS registry number: 113-15-5. Molecular formula: $C_{33}H_{35}N_5O_5$. Molecular weight: 581.66.

The structure of ergotamine is shown in Fig. 2.6.

7.2 Physical data

Description. White powder.

Melting-point. 180 °C.

Spectral properties. UV, IR, and fluorescence spectral data were reviewed by Hofmann (1964). The electron mass spectrum of ergotamine was described by Vokoun and

Řeháček (1975), and the ^1H -NMR spectrum was reported by Pierri *et al.* (1982).

Specific rotation. $[\alpha]_D^{20} -160^\circ$.

7.3 Chemical data

Solubility. Some data on recrystallization, appearance, and solubility were reviewed by Hofmann (1964).

References

- Bezuidenhout SC, Gelderblom WCA, Gorst-Allman CP *et al.* (1988). Structure elucidation of the fumonisins, mycotoxins from *Fusarium moniliforme*. *J Chem Soc Chem Commun*, 11:743–745. doi:10.1039/c39880000743
- Brumley WC, Andrzejewski D, Trucksess EW *et al.* (1982). Negative ion chemical ionization mass spectrometry of trichothecenes. Novel fragmentation under OH⁻ conditions. *Biomed Mass Spectrom*, 9:451–457. doi:10.1002/bms.1200091008
- Budavari S, ed. (1989). *The Merck Index*, 11th ed. Rahway, NJ: Merck & Co.
- Bycroft BW, Hatton JR, Roberts JC (1970). Studies in mycological chemistry. XXV. Experiments directed towards a synthesis of aflatoxin-G₂: synthesis of the coumarinolactone system. *J Chem Soc Perkin 1*, 2:281–284. PMID:5460861
- Castegnaro M, Barek J, Frémy JM *et al.*, eds (1991). *Laboratory Decontamination and Destruction of Carcinogens in Laboratory Wastes: Some Mycotoxins*. Lyon: International Agency for Research on Cancer (IARC Scientific Publications Series, No. 113).
- Castegnaro M, Hunt DC, Sansonne EB *et al.*, eds (1980). *Laboratory Decontamination and Destruction of Aflatoxins B₁, B₂, G₁, G₂ in Laboratory Wastes*. Lyon: International Agency for Research on Cancer (IARC Scientific Publications Series, No. 37).
- Cole RJ, Cox RH (1981). *Handbook of Toxic Fungal Metabolites*. New York: Academic Press.
- Cole RJ, Jarvis BB, Schweikert MA (2003a). Fumonisin, AAL toxins, and related metabolites. In: *Handbook of Secondary Fungal Metabolites, Vol. III*. San Diego: Academic Press, pp. 561–612.
- Cole RJ, Jarvis BB, Schweikert MA (2003b). Ochratoxins and related metabolites. In: *Handbook of Secondary Fungal Metabolites, Vol. III*. San Diego: Academic Press, pp. 613–624.
- Cole RJ, Jarvis BB, Schweikert MA (2003c). Trichothecenes and related metabolites. In: *Handbook of Secondary Fungal Metabolites, Vol. III*. San Diego: Academic Press, pp. 199–324.
- Cole RJ, Schweikert MA (2003). Aflatoxins. In: *Handbook of Secondary Fungal Metabolites, Vol. I*. San Diego: Academic Press, pp. 545–569.
- Gelderblom WCA, Marasas WFO, Vleggaar R *et al.* (1992). Fumonisin: isolation, chemical characterization and biological effects. *Mycopathologia*, 117:11–16. doi:10.1007/BF00497273 PMID:1513367
- Hidy PH, Baldwin RS, Greasham RL *et al.* (1977). Zearalenone and some derivatives: production and biological activities. *Adv Appl Microbiol*, 22:59–82. PMID:412398
- Hofmann A (1964). *Die Mutterkorn-alkaloide*. Stuttgart: Ferdinand Enke Verlag.
- Howard PC, Churchwell MI, Couch LH *et al.* (1998). Formation of *N*-(carboxymethyl) fumonisin B₁ following the reaction of fumonisin B₁ with reducing sugars. *J Agric Food Chem*, 46:3546–3557. doi:10.1021/jf980194q
- Josephs RD, Krska R, MacDonald S *et al.* (2003). Preparation of a calibrant as certified reference material for determination of the *Fusarium* mycotoxin zearalenone. *J AOAC Int*, 86:50–60. PMID:12607740
- Kiermeier F, Kroczeck S (1974). Einfluss der Lösungsmittels auf die Fluoreszenz von Aflatoxin B₁. *Z Lebensm Unters Forsch*, 155:81–84. doi:10.1007/BF01460336
- Kuiper-Goodman T, Scott PM, Watanabe H (1987). Risk assessment of the mycotoxin zearalenone. *Regul Toxicol Pharmacol*, 7:253–306. doi:10.1016/0273-2300(87)90037-7 PMID:2961013
- Lauren DR, Smith WA (2001). Stability of the *Fusarium* mycotoxins nivalenol, deoxynivalenol and zearalenone in ground maize under typical cooking environments. *Food Addit Contam*, 18:1011–1016. doi:10.1080/02652030110052283 PMID:11665729
- Laurent D, Platzer N, Kohler F *et al.* (1989). Macrofusine et micromoniline: deux nouvelles mycotoxines isolées de maïs infesté par *Fusarium moniliforme* Sheld [Macrofusin and micromonilin: two new mycotoxins isolated from corn infested with *Fusarium moniliforme* Sheld]. *Microbiol Aliment Nutr*, 7:9–16.
- Müller HM (1983). A survey of methods of decontaminating mycotoxins. Part II. Chemical methods and reactions with components of feedstuffs. *Übersicht Tierernach*, 11:7–37.
- NTP (2001). *NTP Technical Report on the Toxicology and Carcinogenesis Studies of Fumonisin B₁* (CAS No. 116355-83-0) in F344/N Rats and B6C3F₁ Mice (Feed Studies). Research Triangle Park, NC: National Toxicology Program, U.S. Department of Health and Human Services, National Institutes of Health (NTP Technical Report No. 496; NIH Publication No. 99-3955). Available at http://ntp.niehs.nih.gov/ntp/htdocs/lt_rpts/tr496.pdf.
- Natori S, Sakaki S, Kurata H *et al.* (1970). Chemical and cytotoxicity survey on the production of ochratoxins and penicillic acid by *Aspergillus ochraceus* Wilhelm. *Chem Pharm Bull (Tokyo)*, 18:2259–2268. doi:10.1248/cpb.18.2259 PMID:5494852
- Norred WP, Plattner RD, Dombrink-Kurtzman MA *et al.* (1997). Mycotoxin-induced elevation of free sphingoid bases in precision-cut rat liver slices: specificity of the response and structure-activity relationships. *Toxicol Appl Pharmacol*, 147:63–70. doi:10.1006/taap.1997.8272 PMID:9356308
- O'Neil MJ, Smith A, Heckelman PE, Budavari S, eds. (2001). *The Merck Index*, 13th ed. Whitehouse Station, NJ: Merck & Co.
- O'Neill K, Damoglou AP, Patterson MF (1993). The stability of deoxynivalenol and 3-acetyl deoxynivalenol to gamma irradiation. *Food Addit Contam*, 10:209–215. doi:10.1080/02652039309374143 PMID:8314397
- Pierri L, Pitman IH, Rae ID *et al.* (1982). Conformational analysis of the ergot alkaloids ergotamine and ergotaminine. *J Med Chem*, 25:937–942. doi:10.1021/jm00350a010 PMID:7120281
- Plattner RD, Norred WP, Bacon CW *et al.* (1990). A method of detection of fumonisins in corn samples associated with field cases of equine leukoencephalomalacia. *Mycologia*, 82:698–702. doi:10.2307/3760156
- Pohland AE, Schuller PL, Steyn PS, Van Egmond HP (1982). Physico-chemical data for selected mycotoxins. *Pure Appl Chem*, 54:2219–2284. doi:10.1351/pac198254112219
- Robertson JA, Pons WA (1968). Solid state fluorescence emission of aflatoxin on silica gel. *J Assoc Off Anal Chem*, 51:1190–1192.
- Ryu D, Hanna MA, Bullerman LB (1999). Stability of zearalenone during extrusion of corn grits. *J Food Prot*, 62:1482–1484. PMID:10606157
- Ryu D, Hanna MA, Eskridge KM, Bullerman LB (2003). Heat stability of zearalenone in an aqueous buffered model system. *J Agric Food Chem*, 51:1746–1748. doi:10.1021/jf0210021 PMID:12617617
- Savard ME, Blackwell BA (1994). Spectral characteristics of secondary metabolites from *Fusarium* fungi. In: Miller JD, Trenholm HL, eds. *Mycotoxins in Grain: Compounds Other than Aflatoxin*. St Paul, MN: Eagan Press, pp. 59–260.
- Steyn PS, Holzapfel CW (1967). The synthesis of ochratoxins A and B metabolites of *Aspergillus ochraceus* Wilh. *Tetrahedron*, 23:4449–4461. doi:10.1016/S0040-4020(01)88843-8 PMID:6077765
- Stubblefield RD, Shotwell OL, Shannon GM *et al.* (1970). A new metabolite from *Aspergillus parasiticus*. *J Agric Food Chem*, 18:391–393. doi:10.1021/jf60169a025 PMID:5487091
- Subirade I (1996). Fate of ochratoxin A during breadmaking. *Food Addit Contam*, 13 Suppl:25–26. PMID:8972345

- Uwaifo AO, Emerole GO, Bassir O (1977). Comparative study of the fluorescent characteristics of solutions of aflatoxins and palmotoxins in chloroform. *J Agric Food Chem*, 25. doi:10.1021/jf60213a021 PMID:893818
- Van der Merwe KJ, Steyn PS, Fourie L (1965a). Mycotoxins. II. The constitution of ochratoxins A, B, and C, metabolites of *Aspergillus ochraceus* Wilh. *J Chem Soc Perkin 1*, 7083–7088. doi:10.1039/jr9650007083 PMID:5892024
- Van der Merwe KJ, Steyn PS, Fourie L *et al.* (1965b). Ochratoxin A, a toxic metabolite produced by *Aspergillus ochraceus* Wilh. *Nature*, 205:1112–1113. doi:10.1038/2051112a0 PMID:5833211
- Van der Stegen GH, Essens PJ, van der Lijn J (2001). Effect of roasting conditions on reduction of ochratoxin A in coffee. *J Agric Food Chem*, 49:4713–4715. doi:10.1021/jf0105586 PMID:11600012
- Visconti A, Doko MB, Bottalico C *et al.* (1994). Stability of fumonisins (FB₁ and FB₂) in solution. *Food Addit Contam*, 11:427–431. doi:10.1080/02652039409374244 PMID:7958112
- Vokoun J, Řeháček Z (1975). Mass spectra of ergot peptide alkaloids. *Collect Czech Chem Commun*, 40:1731–1737.
- WHO (2000). *Environmental Health Criteria 219: Fumonisin B₁*. Marasas WFO, Miller JD, Riley RT, Visconti A, eds. Geneva: United Nations Environment Programme, International Labour Organization, World Health Organization. Available at http://libdoc.who.int/ehc/WHO_EHC_219.pdf.
- Widestrand J, Pettersson H (2001). Effect of time, temperature and solvent on the stability of T-2 toxin, HT-2 toxin, deoxynivalenol and nivalenol calibrants. *Food Addit Contam*, 18:987–992. doi:10.1080/02652030110050168 PMID:11665740
- Wogan GN (1966). Chemical nature and biological effects of the aflatoxins. *Bacteriol Rev*, 30:460–470. PMID:5327461
- Wolf CE, Bullerman LB (1998). Heat and pH alter the concentration of deoxynivalenol in an aqueous environment. *J Food Prot*, 61:365–367. PMID:9708313
- Wolf-Hall CE, Hanna MA, Bullerman LB (1999). Stability of deoxynivalenol in heat-treated foods. *J Food Prot*, 62:962–964. PMID:10456755